

Fraction A contains high molecular materials, mainly proteins and s-RNA. The other fractions (B, C, D, E) are of low molecular nature; all pass through a dialysation membrane. The chemical analysis and the maximum of the UV-absorption of these fractions are shown in the Table. Fraction B includes substances containing sugars, aminosugars, sugar phosphates and some of the amino

Analysis of low molecular fractions of rat liver

Fraction	B	C	D	E
Ninhydrin colour*	157.5	14.9	42.5	117.0
Ninhydrin colour after hydrolysis	202.5	390.0	960.0	860.0
Phosphorus %	12.45	0.184	0.914	0.38
Carbohydrate %	33.7	0.768	0.557	0.526
Hexosamine after hydrolysis %	0.586	—	—	—
Maximum UV-absorption m μ	265	260	255	275

* Ninhydrin colour expressed as μ g of leucin/mg

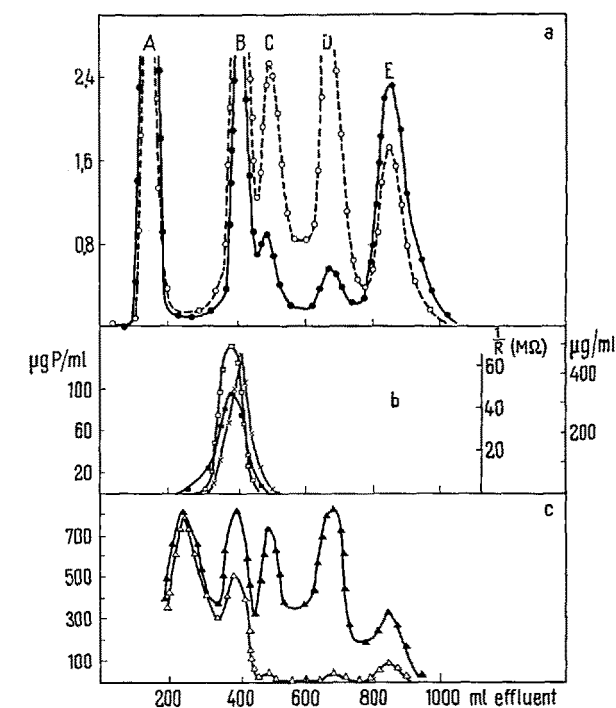
acids and low peptides. Amino acids also eluate between the peak A and B. Materials containing sugar are eluted from the column in the ascending part of peak B. The fractions C, D, E contain mainly peptides, nucleotides and nucleopeptides.

All low molecular fractions are heterogeneous in the electric field. Lyophilized fractions were separated by preparative electrophoresis on paper in 0.1 M ammonium acetate buffer pH 5.9. All chromatographic fractions contain compounds with UV-absorption and are ninhydrin positive⁸.

Zusammenfassung. Es werden einige niedrigmolekulare Stoffe aus Rattenleberextrakt nach der Sephadex G25-Fraktionierung beschrieben. Die Lokalisierung von Zucker, Aminosäuren, Peptiden, Nukleopeptiden und verwandter Stoffe wurde im Elutionsdiagramm vorgenommen.

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Elution diagram of rat liver extract on the column Sephadex G25. ○—○ O.D. at 260 m μ ; ●—● O.D. at 280 m μ ; ×—× conductivity; □—□ carbohydrate (Bial) as μ g/ml; ■—■ phosphorus μ g/ml; △—△ ninhydrin colour; ▲—▲ ninhydrin colour after hydrolysis expressed as μ g of leucin/ml.

Spontaneous Leukaemia in a Sprague-Dawley Rat

Spontaneous leukaemia in rats is remarkably rare. Few reports are available since 1936 when WILENS and SPROUL observed the first cases¹⁻³. A spontaneous myeloid chloroleukaemia is described here since (1) no incidence has been reported yet in Sprague-Dawley rats, and (2) the disease was accompanied by some intriguing renal tubular changes. The nature of these changes and their relation to the leukaemic process is not clear.

The rat was a 13-month untreated female of the above mentioned strain.

Hematology: WBC, 77000; RBC, 5.2 millions; Hgb, 13.3 g. The peripheral blood smear (Wright's stain) showed evidence of leukaemic invasion by immature cells of the granulocyte series (Figure 1). Two types were prominent: (1) large cells with large rounded nuclei and very little basophilic cytoplasm containing a small amount of azurophil granulation, and (2) large cells with rounded or polygonal nuclei and a greater amount of polychromatophil or acidophil cytoplasm containing numerous neutrophil granules. The differential count revealed that 70% of the white blood cells were Type 1

(promyelocyte) and Type 2 (myelocyte) cells with a number of atypical forms.

Significant autopsy findings included: *spleen*—enlarged to ten times normal size, solid, greyish coloured; *lymph nodes*—markedly enlarged, characteristic light green colour; and femoral *bone marrow*—similar green colour.

Microscopically the cellular infiltration was most conspicuous in the R.E. organs. The liver showed an extreme degree of infiltration of the periportal, perivascular spaces and sinusoids by immature myeloid elements together with considerable liver cell damage.

¹ S. L. WILENS and E. E. SPROUL, Amer. J. Path. 12, 249 (1936).

² C. H. OBERLING, M. and P. GUERIN, Bull. Assoc. fran \acute{c} ais. l'etude Cancer 28, 214 (1939).

³ G. ROUSSY, M. and P. GUERIN, Bull. Assoc. fran \acute{c} ais. l'etude Cancer 30, 29 (1942).

⁴ H. D. KESTEN, unpublished data (1946) from H. SHAY et al., Blood 7, 613 (1952).

⁵ R. IGLESIAS and E. MARDONES, Proc. Amer. Assoc. Cancer Res. 2, 121 (1956).

⁶ W. F. DUNNING and M. R. CURTIS, J. Nat. Cancer Inst. 19, 845 (1957).

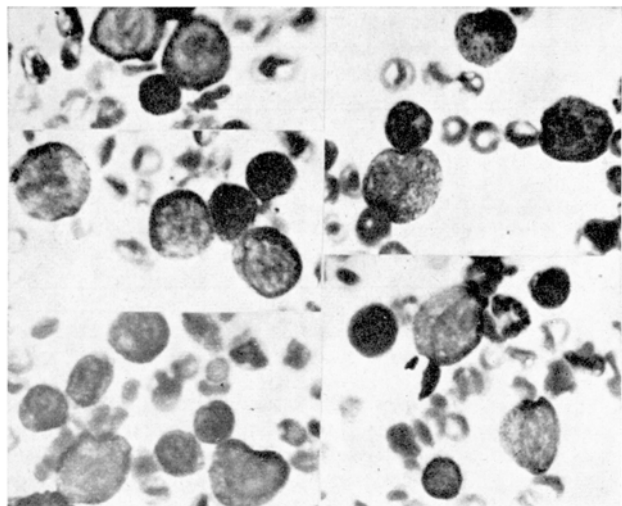


Fig. 1. Peripheral blood smear (Wright's stain).

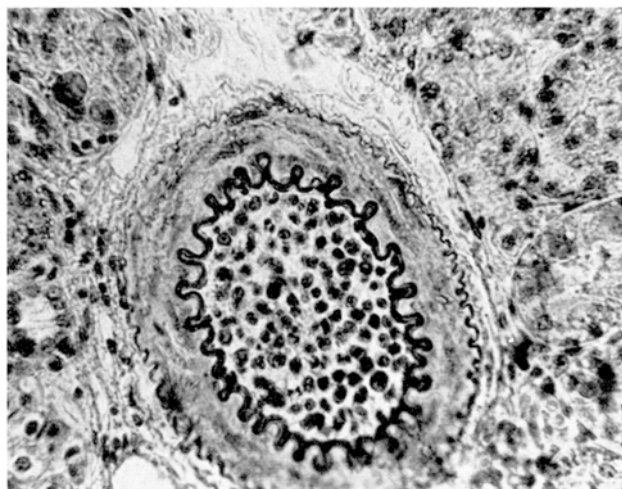


Fig. 2. Renal cortical artery (frozen, Oil red O-Hematoxylin, 310 \times).

Spleen and lymph nodes—follicular structure was almost completely obliterated and replaced, also infiltration of the perinodal fat was evident. Bone marrow—there was replacement of the normal cell population by early myeloid cells with few erythrogenic foci, and far less fat was present than in normal bone marrow.

In contrast with other organs the kidney did not show any substantial infiltration, however, most of the larger blood vessels of the cortex contained a great number of leukaemic cells in their lumens (Figure 2). In the outer



Fig. 3. Renal tubules (PAS, oil immersion).

zone of the cortex a pronounced vacuolar change with cytoplasmic droplet formation was seen (Figure 3). Sharply contoured homogeneous droplets and globules of various sizes (the largest exceeding the diameter of the tubule cell nucleus) appeared to be in a process of fusion and aggregation. Staining characteristics of the globules were as follows: eosinophilic, Oil red O negative; hemoglobin negative (Dunn-Thompson), strongly PAS positive, violet with Weigert fibrin stain and dark blue with Mallory's PTAH method.

The hematology and pathological picture in our chloroleukaemic rat were basically identical with those cases reported previously, also the green colour was mainly localized (as in HALL and KNOCKE's chloroleukaemic mouse) in the lymph nodes⁷. The peculiar tubular droplet change has been mentioned only briefly by two previous groups^{1,3}, who suspected a rather specific link between this tubular alteration and leukaemia in rats. In our case the physical and staining properties of the droplets indicated deposition of a substance of essentially protein character but the exact nature, origin and significance require further study.

Résumé. Description d'un cas de chloroleucémie myéloïde chez un rat de souche Sprague-Dawley, présentant quelques modifications peu communes des tubes renaux.

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Research Department, Frank W. Horner Limited, Montreal, Quebec (Canada), August 13, 1962.

⁷ J. W. HALL and F. J. KNOCKE, Amer. J. Path. 14, 214 (1938).

Dimorphic Sperms of *Rhinopoma kinneari* (Chiroptera)

Many publications have appeared in the past on the dimorphism of the sperms in mammals. The morphological differences in the two populations of sperms have been found either in the size and the shape of the nuclei¹, or in the relative size differences of the centrally located

heterochromosomes², or in the head lengths of the dimorphic sperms³.

Stained and unstained preparations of the freshly ejected sperms of *Rhinopoma kinneari* (Rhinomatidae,

¹ L. B. SHETTLES, Fertility and Sterility 12, 20 (1961).

² L. B. SHETTLES, Nature 187, 254 (1960).

³ A. S. PARKES, Quart. J. micr. Sci. 67, 617 (1923).